A NEW AMINO ACID, NICOTIANAMINE, FROM TOBACCO LEAVES

M. Noma, M. Noguchi, and E. Tamaki

(Central Research Institute, Japan Monopoly Corporation)

Nishi-shinagawa, Shinagawa-ku, Tokyo, Japan.

(Received in Japan 29 March 1971; received in UK for publication 30 April 1971) In the course of our studies of nitrogen metabolism in tobacco plants, a new amino acid was isolated from, Nicotiana tabacum L. "Hicks".

We wish to report here the isolation and the structure elucidation of the amino acid tentatively named "nicotianamine" (represented as (I) in the following text).

Green tabacco leaves (70 Kg) were extracted 3 times with 70 % aqueous MeOH. The aqueous extract obtained by evaporating MeOH in vacuo was treated with charcoal, and the resultant clear solution was further treated with Dowex 50W-X4 resin. The cationic constituents thus obtained were subjected to displacement chromatography with a set of five columns of Dowex 50W-X4 resin according to the method of Partridge et al¹⁾. Fractions containing (I) were then submitted to cellulose column chromatography with n-BuOH: acetic acid: water (4:1:5) as solvent to yield almost pure compound(I). Recrystallization from water twice afforded pure 209 mg of (I).

This compound (I), $[\mathbf{O}_{D}]_{D}^{23} = -60.5$ (C=2.7, in water), with $C_{12}H_{23}N_{3}O_{7}$ by elementary analysis, gradually darkened in colour above 240° C without melting, is labile to mineral acid and a test for \mathbf{O}_{1} -amino acid²) is positive. The elution peak with an amino acid analyzer in the usual physiological fluid analysis is overlapped with those of asparagine and glutamine.

In the NMR spectrum in D_2O (Fig. I), the following signals are observed: 4H(m), $\delta 2.2$; 2H(m), $\delta 2.6$; 2H(t), $\delta 3.2$; 2H(t), $\delta 3.4$; 4H(m), $\delta 4.0$. No signal assigned to methyl group is detected.

In the mass spectra of the methyl (Fig. 3) and the ethyl ester, parent peaks were observed at m/e 345 ($C_{15}H_{27}N_3O_6$) and m/e 387, base peaks at m/e 128 ($C_6H_{10}NO_2$) and m/e 142 respectively, indicating that (I) exists as a monohydrate and has three carboxyl groups. Since it absorbs no hydrogen atom on catalytic hydrogenation on platinum black, one ring system must be included in the molecule.

In a dcutcro labelling experiment³⁾, the parent peak of the methyl ester was shifted to m/e 348, but the base peak was not. These indicate that the methyl ester has three active hydrogen

atoms attached to basic nitrogen atoms and the nitrogen atom in the base peak has none of such hydrogen atoms. From the difference of 14 mass units between the base peak of the methyl ester and that of the ethyl ester, two oxygen atoms in the former are suggested to be attributed to a On the other hand, $KMnO_{L}$ oxidation of (I) (1.1 μ mole), yielded aspartic carbomethoxy group. acid (0.4 µmole), g-alanine (0.09 µmole) and azetidine-2-carboxylic acid (0.2 µmole). The presence of azetidine-2-carboxylic acid moiety in (I) was further supported by the comparison of the NMR spectrumn of (I) with that of azetidine-2-carboxylic acid. In the latter, two 2H multiplets assigned to CH₂ at 3 and 4 position of the azetidine ring are observed at §2.6 and §4.0 respectively. Coupling pattern of the signal at $\delta 2.6$ is similar to that of (I) at $\delta 2.6$. Thus, 4H multiplet at 4.0 in (I) implies the 2H multiplet attributed to 4 position of the azetidine ring. In the both compounds the signal due to the proton attached to 2-position of carbon atom in the azetidine ring is hidden by the strong signal due to HDO at $\S4.7$. Considering the above results, and admitting that the base peak in the mass spectrum of the methyl ester is generated by primary fragmentation, the structure (A) is given for

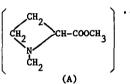
the base peak. The existence of the m/e 114 $(C_L H_R NO_2)$

m/e 142 $(C_7H_{12}NO_2)$ and m/e 156 $(C_8H_{14}NO_2)$ peaks in addition to m/e 128, suggests that at least 3 CH_2 units combine linearly to the nitrogen atom of the azetidine ring.

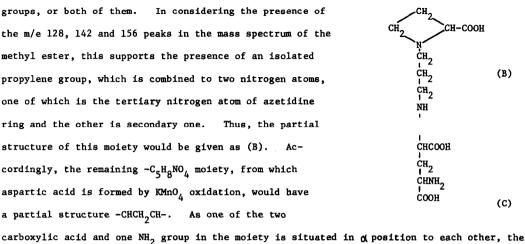
By KMnO_4 oxidation of azetidine-2-carboxylic acid (0.17 µmole), aspartic acid and β -alanine were obtained, yielding 0.005 µmole and 0.03 µmole respectively. Comparing these results with the yields of the oxidation products of (I), most of aspartic acid formed in the oxidation of (I) is assumed not to be derived from azetidine-2-carboxylic acid molety, but from the other molety which is easily oxidized to aspartic acid.

The U.V. spectrum of the DNP derivative of (I) has max at 360 mm with ξ = 1.6 x 10⁴, showing that one NH₂ group exists in (I). Thus, the three nitrogen atoms in (I) are supposed to exist in one primary, one secondary and one tertiary amine. KMnO₄ oxidation of the DNP derivative afforded DNP-aspartic acid. These indicate that the NH₂ group in (I) exists in the moiety which yields aspartic acid by KMnO₄ oxidation.

By the double resonance experiment irradiated at §2.2, triplets at §3.4 and §3.6 changed to two 2H singlets and two 1H singlets appeared near §4.0 remaining the unchanged multiplet signals due to the two protons in 4 position of the azetidine ring. This indicates that either two -CH₂CH₂CH₋, or one-CH₂CH₂CH₂- and one -CHCH₂CH- groups are present in the molecule where the pro-



tons attached to the both terminal carbons are deshielded by adjacent nitrogen atoms, carboxyl



partial structure (C) is given for this moiety.

In the high resolution mass spectrum of the methyl ester, the existence of the m/e 257 $(C_{12}H_{21}N_2O_4, M-CHNH_2COOCH_3), m/e 243 (C_{11}H_{19}N_2O_4, M-CH_2CHNH_2COOCH_3), m/e 171 (C_8H_{15}N_2O_2, M-CHNH_2COOCH_3), m/e 171 (C_8H_{15}N_2O_2, M-CHNH_2COOCH_3), m/e 171 (C_8H_{15}N_2O_2, M-CHNH_2COOCH_3), m/e 171 (C_8H_{15}N_2O_3, M-CHNH_2COOCH_3), m/e 171 (C_8H_{15}N_2O_3), m/e 1$ CHCOOCH₃CH₂NH₂COOCH₃) and m/e 174 (C₂H₁₂NO₄, CHCOOCH₃CH₂CHNH₂COOCH₃) peaks established the structure of (I) as 1-[3'-(Y-amino-d,f-dicarboxy propylamino)-propyl]-azetidine-2-carboxylic acid.

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Reference

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(B)

(C)

